

***Metrology for monitoring endocrine  
disrupting compounds under the Water  
Framework Directive\_EDC-WFD***

# PRESENTATION/ CONTEXT

# THE CONSORTIUM

- 8 partners / 6 European countries
- 5 NMI/DI, one DI operating outside of its designation, one academic research laboratory, one research institute
- Consortium brings together **scientific excellence** in research institutes and experience in **ultra-trace measurements of micropollutants**
- **Balance of expertise:** development and certification of RM, proficiency tests / interlaboratory comparison design, method development and validation, standardisation



**Start date: 1<sup>st</sup> September 2019**  
**Duration: 36+6 months**  
**Budget: 800K€**

# THE PROJECT

## AIMS:

- ❖ Address the standardization lack for harmonised measurement methods for estrogens in whole water
- ❖ Ensure that measurements of estrogens are traceable, well defined, meet the requirements of the WFD, and thus are comparable across Europe (and worldwide)

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- Chief Stakeholder: Ulrich BORCHERS Chairman of CEN/TC 230
  - Chief Stakeholder and DIN secretary (Andreas Paetz) are kept inform of the progress

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- Support from AFNOR (Arnaud Gaudrier: Secretary of SC «Water Quality»)

# CONTEXT

- **Estrogens:** group of chemicals of similar structure mainly responsible for female sexual development and reproduction.
- In water ecosystem:
  - **Pseudo-ubiquitous** and occur at **ultra-trace level** ( $\ll \text{ng L}^{-1}$  to  $\text{tens ng L}^{-1}$ )
  - Level at which they can have effects in natural species
    - **Threat for biodiversity**



*Included in the First Watch List of the Water Framework Directive WFD*



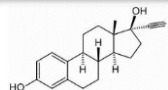
*Under review to become priority substances (PS) of the WFD*

- **No EN or ISO standard for MS-based methods** currently available or in progress
- (Accredited) testing laboratories **develop and validate in-house methods** according to internal criteria
- Most of (accredited) testing laboratories **failed to achieve the very low LOQ** to enable monitoring of estrogens at relevant level
- **Metrological endpoints** have been highlighted of particular importance if effect-based method (EBM) results are to be used in a regulatory context
- **Lack/absence of reference materials and ad'hoc proficiency tests**

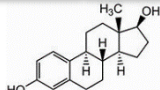
# THE PROJECT

## Targeted substances

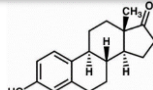
Candidate PS



17α-ethinylestradiol

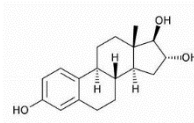


17β-estradiol



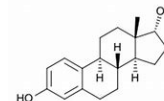
Estrone

+



Estriol

Stability + cross reactivity of EBM



17α-estradiol

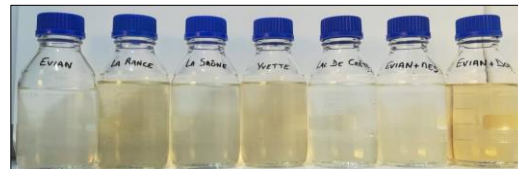
Specificity of MS-based method + cross reactivity of EBM

## Targeted level of concentrations (EQS)

Inland waters	PNEC beginning of the project ( $\mu\text{g L}^{-1}$ )	EQS SCHEER final decision march 2022 ( $\mu\text{g L}^{-1}$ )
17α-ethinylestradiol	0.000035	0.000017
17β-estradiol	0.0004	0.00018
Estrone	3.6	0.00036

## Target Matrix

- Inland freshwaters: surface water and ground water (opt. drinking water)
- Representative of European inland waters panel



# AIM

Natural and pharmaceutical estrogens are key Endocrine Disrupting Chemicals (EDC) which are monitored differently depending on the country, and for which standardised reference methods are currently not available.

⇒ Main Objective: Develop reliable and harmonized measurement methods for estrogens, to comply with the WFD Directive requirements

⇒ Outcomes: to be disseminated to CEN/ TC 230 and ISO/ TC 147 to be fed into the documentary standards they develop

# OBJECTIVES

- The overall objective of this project is to **develop traceable measurement methods** for endocrine disrupting chemicals, with a specific focus **on three estrogens of the first watch list: 17-beta-estradiol (17 $\beta$ E2), 17 alpha ethinylestradiol (17 $\alpha$ EE2), and estrone (E1))**
- Estrogens 17-alpha-estradiol (17 $\alpha$ E2) and estriol (E3) will be included to demonstrate the **reliability of the developed methods**
  - ⇒ **to support the requirements of Directive 2013/39/EC, Directive 2009/90/EC and Commission Implementation Decision (EU) 2018/840,**
  - ⇒ **improve the comparability and compatibility of measurement results within Europe**



■ The specific objectives of the project are to:

- 1. Optimize and validate traceable aqueous reference Mass Spectrometry-based methods for the analysis of 5 estrogenic compounds** prioritizing  $17\beta\text{E}2$ ,  $17\alpha\text{EE}2$ , and E1 in **whole water samples** at environmental quality standard (EQS) levels. **Methods will have limit of quantification (LOQ) not exceeding 30% EQS with a measurement uncertainty of  $\leq 50\%$  at EQS**
- 2. Develop production methods for aqueous reference materials (RM)**, which are as close as possible to real water samples, with proven homogeneity, short- and long-term stability
- 3. Improve the comparability of estrogen measurements with selected Effect-Based Methods (EBM)** in whole water samples at EQS level. Methods will have been correctly calibrated and information on uncertainty will be provided
- 4. Organize and perform an interlaboratory comparison (ILC)** to demonstrate the performance of the developed methods using the reference material (RM) for the selected estrogen substances
- 5. Contribute to the work of key European and international standardization organizations e.g. CEN TC 230 and ISO TC 147**



Ultratraces estrogens  
measurements  $< 0,1\text{ng/L}$   
In whole water

⇒ New dimension

**Every detail counts**

# WHAT DO WE NEED TO MEASURE? WHAT ARE WE MEASURING?

# WHOLE WATER MEASUREMENTS WHAT ARE WE TALKING ABOUT ?

- What we have in essence developed and validated are whole water analysis methods
- Also **Water Framework Directive** requires to analyze whole water
- Whole water is composed of **liquid phase** and **particulate phase**. **Analytes can exist in both phases.**
- When analyzing whole water, one collects, concentrates, extracts both liquid and particulate phases so that **analytes in both phases are captured and analyzed.**

# WHOLE WATER MEASUREMENTS WHAT ARE WE TALKING ABOUT ?

- The concentration result obtained is **a sum of the analyte concentration in both phases.**
- Sample preparation is started by adding internal standards and by letting them to **equilibrate between liquid and particulate phases** for at least 12 hours.
- This improves the probability that results are accurate even if analyte is not fully recovered from the particulate phase in sample extraction.

# SAMPLING

# SAMPLING

- ❑ Comply with general principles of ISO 5667 guidelines
- ❑ Implement general principles for ultratrace analysis
- ❑ No specific risk or source of contamination was identified during field sampling in the project

- ❑ Recommendations:



- use glass bottle and prevent photodegradation of analytes (amber, green bottles, alumina foil, ...)
- Avoid extra steps, intermediate containers in order to minimize risk of cross-contamination and adsorptive losses
- Sample container cleaning procedures should be implemented: For example calcination of glassware, rinsing containers with solvents



Avoid plastic containers in case of EBMs

## Illustration of adsorption phenomena

Table 2. Relative mass lost over 24 h and first-order sorption rates for selected r

Material	%E2†	%EE2	%E1
Type 304 stainless steel	24.9 ± 5.5	53.4 ± 2.0	52.7 ± 2.4
Type 316 stainless steel	30.5 ± 1.8	56.1 ± 3.7	54.3 ± 3.6
Glass (culture tubes)	1.0 ± 0.7	0.7 ± 0.6	0.6 ± 0.4
PolyCarbonate	8.7 ± 5.1	51.2 ± 8.3	44.1 ± 6.8
PVC‡	4.5 ± 2.5	5.0 ± 2.1	7.7 ± 4.5
Teflon	2.3 ± 1.3	4.2 ± 0.6	2.2 ± 0.8
Autoclaved			
Type 304 stainless steel	2.3 ± 7.0	33.7 ± 7.6	28.4 ± 6.1
Type 316 stainless steel	6.8 ± 17.7	33.2 ± 6.2	20.9 ± 5.8
Glass (culture tubes)	-4.9 ± 0.6	-0.2 ± 1.2	0.5 ± 0.8

➤ Minimum losses with glass container and materials

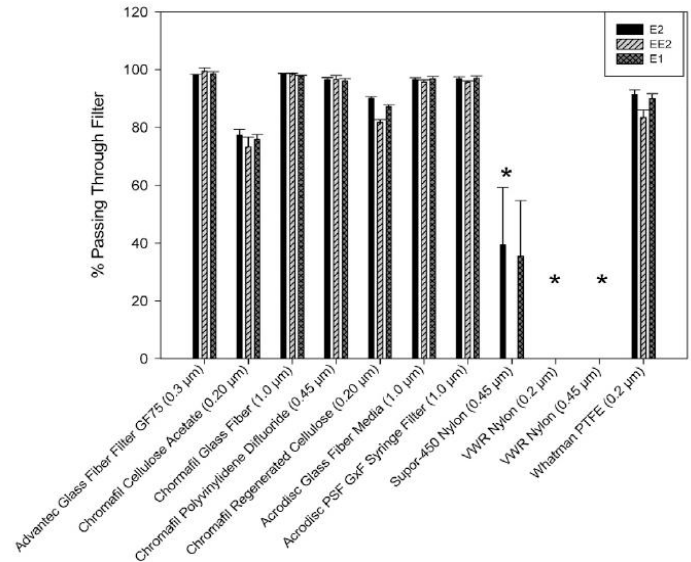


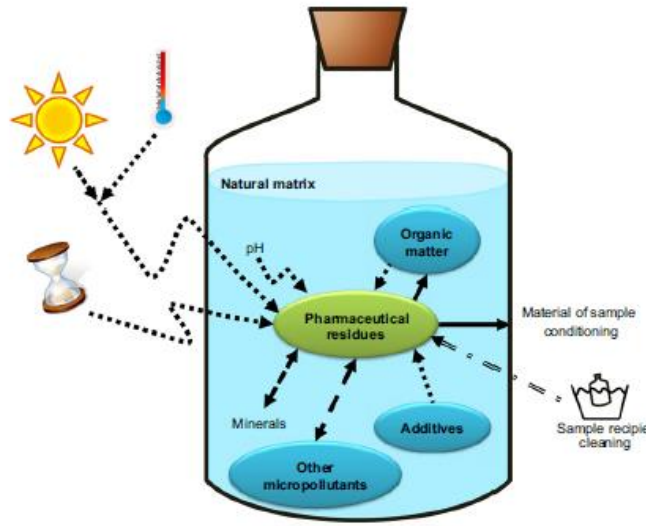
Fig. 3. Percent passing through different filter types. \*Significant differences ( $p < 0.05$ ) between the percent passing and the initial concentration. Error bars represent 1 SD of the mean for triplicate samples. E1, estrone; E2, 17 $\beta$ -estradiol; EE2, 17 $\alpha$ -ethynylestradiol.



# STABILITY

# Stability

## Main drivers



- ..... → Degradation (biodegradation, biotodegradation, photodegradation, hydrolysis)
- Adsorption
- - - - - → Formation of complex, oxidation, reduction
- : = ⇒ Cross contamination

Fig. 1. Sources and processes (possibly) affecting the stability of PPs in samples before analysis.

Numerous reviews and publications


⇒ Not all in agreement

- Methodological strategy
- Acceptance criteria
- Missing key information

Mompelat et al. (2013)

# Stability

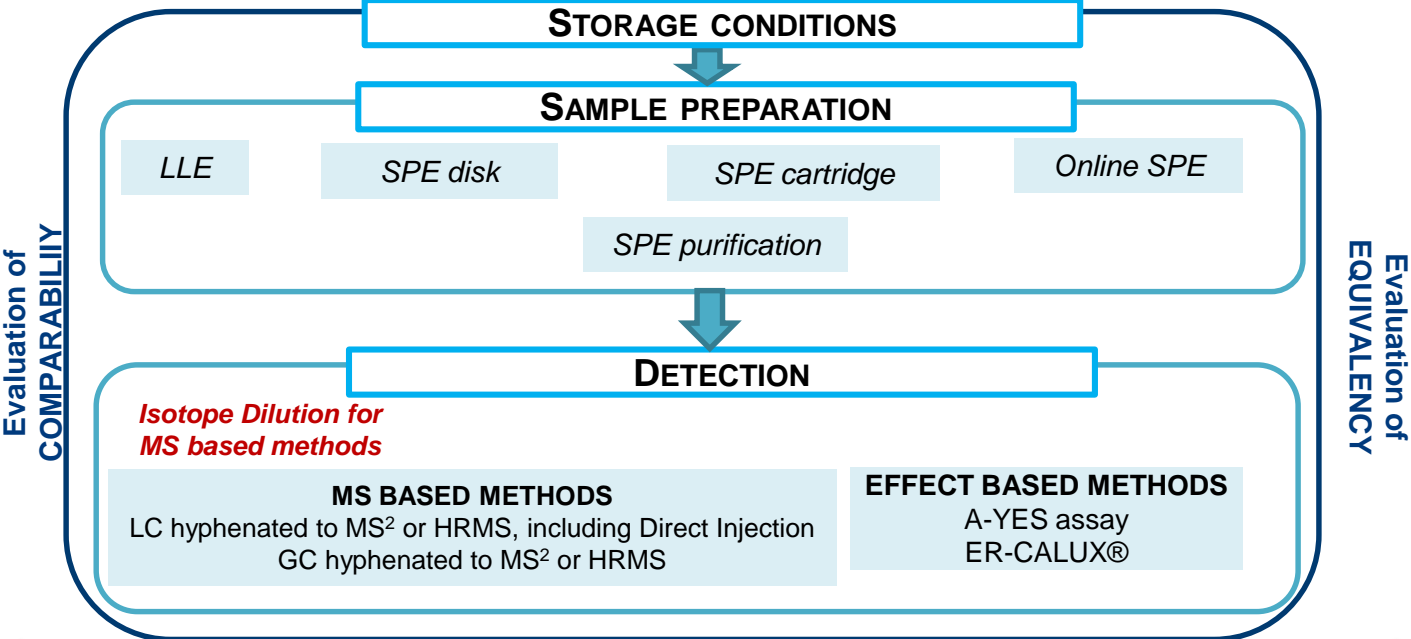
## □ Recommendations:

- 
- Glassware (coloured) shall be preferred over other type of containers if very low LOQs ( $\ll 0,1\text{ng/L}$ ) are targeted, to minimize the risk of analyte losses due to adsorption
  - Storage at  $4^{\circ}\text{C}$  is OK for 14 days for synthetic samples of low and high complexity
  - Storage time is expected to be shorter for complex natural samples. Addition of 1% MeOH is recommended to avoid biodegradation because it has not shown negative effects on recoveries
  - Storing samples at  $-20^{\circ}\text{C}$  is possible but it may increase risk of clogging during sample extraction if SPE cartridges are used for extraction.



Plastic container shall be avoided if LOQ  $\ll 0,1\text{ng/L}$  targeted

# METHOD OPTIMIZATION



Comparison of sample preparation techniques for estrogens in whole water & recommendations on the most appropriate ones

Most promising MS-based method(s) for the measurement of selected estrogens in whole water samples compatible with the requirements of the QA/QC Directive (LOQ ≤ 30% EQS with U(k=2) ≤ 50% at EQS)

Recommendations to improve the comparability of estrogens measurements with the set of selected EBMs in whole water samples at EQS levels

