

**Sample preparation  
strategies for estrogens  
with regards to the EU-  
WFD**

## ***C1 WP1: Optimisation and evaluation of sample preparation methods***

The aim of this work package is to evaluate and optimise the sample preparation for a set of various whole water samples including freshwater (surface and ground water) and a synthetic real matrix water which consists of distinct ingredients e.g. dissolved organic compound (DOC) (humic substances) and suspended particulate matter (SPM).

### **B1.c List of deliverables**

<b>Relevant objective (Activity delivering the deliverable)</b>	<b>Deliverable number</b>	<b>Deliverable description</b>	<b>Deliverable type</b>	<b>Partners (Lead in bold)</b>	<b>Delivery date</b>
Objective 2 (A1.3.4)	D1	Report comparing sample preparation techniques for estrogens partitioning in whole water and recommendations on the most appropriate methods	Report Recommendations	<b>BAM</b> , JSI, LNE, TUBITAK, SYKE, UBX	Apr 2021 (M20)



Oct. 2021

## Needs for sample preparation

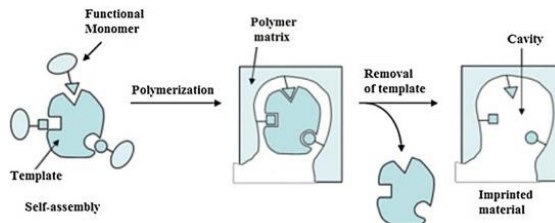
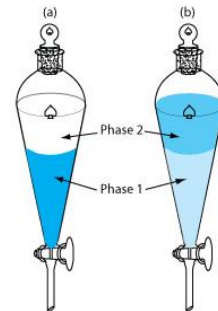
In general, a compromise is necessary, which addresses the whole water sample, a reasonable preconcentration factor, and the robustness of the method:

- Preconcentration of target analytes due to low EQS given by the EU-WFD
- Separation of matrix components from target analytes to prevent matrix effects on the chromatographic separation e.g., decreasing separation efficiency of the chromatographic column and enhancing or decreasing signal intensity (ion suppression in ESI source)
- Avoiding contamination of sample inlets (GC liner contamination by matrix components)

A preselection of feasible and applicable techniques was done by each project partner according to their practical knowledge and available instrumentation

# Overview on preconcentration procedures

- Liquid-liquid extraction (LLE)
- Solid phase extraction (SPE) columns (off- and online)
- SPE disks
- Mi-SPE (molecular imprinted polymers)

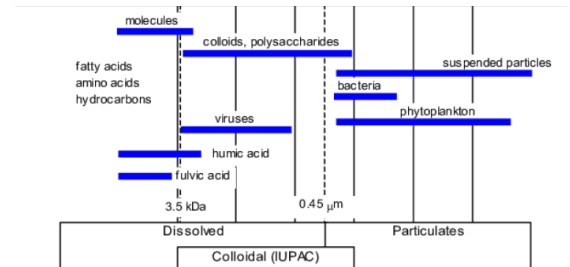


# Whole water matrix simulation

- Evian water (bottled in glass, if available)



- Addition of SPM (0 - 50 mg/l)
- Addition of DOC (0 - 14 mg/l)
- Addition of low ng/L of estrogens
- Internal standards



1. Continuum of particles, colloids and dissolved organic carbon in natural waters (Aiken and Leenheer, 1993)

## Liquid-liquid extraction (LLE)

- High volumes of organic extraction solvents needed (30 – 50 mL; green chemistry)
- Limited selectivity (non-miscible organic solvents only)
- “Batch extraction” (must be repeated with portions of fresh extraction solvent)
- Not automatable (depending on who does the extraction, no extractions in parallel): not suitable for routine laboratories
- Batch extracts must be combined and evaporated



**Not applicable in routine lab!**

# Liquid-liquid extraction (LLE)

Typical procedure:



500 mL water sample

- 3 x extraction with 30 mL DCM
- Extraction time 1 min/set 10 min



Combining organic layers

- Evaporation to dryness
- Reconstitution with 1 mL ACN

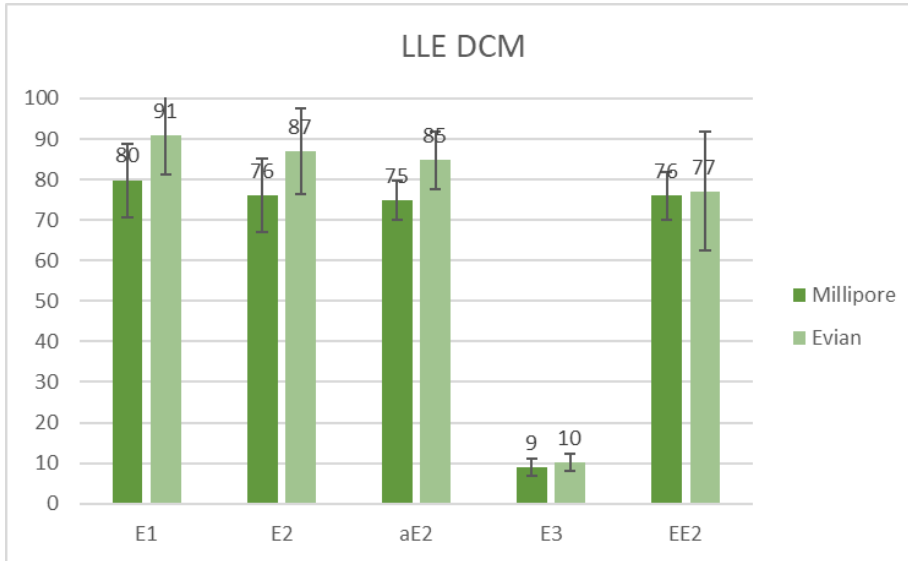


Analysis

- The ACN extract is ready for analysis



# Liquid-liquid extraction (LLE)



- Sufficient absolute recoveries for E1, E2, alphaE2 and EE2 but poor for E3, independently from the type of water matrix



## Solid phase extraction (SPE) off- or online

- e.g., HLB (Hydrophilic-Lipophilic Balanced) 6 or 3 mL / filled with 150 or 60 mg sorbent material. Other materials are also feasible ( $C_{18}$ ,  $C_{18}$  eq, ...)
- With a content of less than  $50 \text{ mg L}^{-1}$  SPM no clogging can be observed (it strongly depends on the ration of SPM to sample volume)
- Preconcentration depends not only on the target analytes, but also on enrichment of DOC content (evaluated by measuring the DOC of the sample extract)
- Problems of ion suppression when using LC-MS/MS (in most cases decrease of the signal intensity, known effect in the ESI source)



OLISIS<sup>®</sup>  
SAMPLE EXTRACTION PRODUCTS



## A typical SPE procedure



### 1000 mL water sample

- Spiked with IS-mix at desired level
- 15 min on horizontal shaker
- Oasis HLB 3 mL/100mg



### Conditioning / sample load

- Conditioning with 10 mL ACN
- Conditioning with 15 mL H<sub>2</sub>O
- Load 1000 mL water sample 20 mL/min



### Washing step and elution

- Washing cartridge with 10 mL H<sub>2</sub>O
- Dry cartridge with N<sub>2</sub> for 1 min
- Collect 10 mL fraction into sample tube using ACN



### Concentration and analysis

- Concentrate organic fraction to 1 mL (TurboVap)
- Extract is ready for analysis by LC-MS/MS



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## Solid phase extraction (SPE) off- or online

- Different matrix compositions were used:
  - **Low complexity matrix** (Evian water with low DOC and without SPM)
  - **High complexity matrix** (Evian water with moderate DOC and with SPM)
- Relative and absolute recovery rates were determined to compare the different experiments with regards to their performance (applicability in routine analysis)

# Solid phase extraction (SPE) off- or online

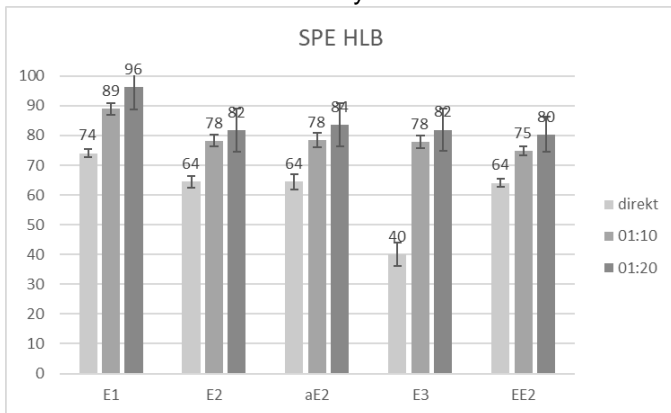
Low complex matrix (Evian water with low DOC and without SPM)

Extraction method  Parameter	Waters Prime HLB size 3cc/60mg, Evian water (500ml) + DOC (7 mg/L) and estrogens at 5 ng/L level			
	Absolute recovery (%)	RSD (%)	Relative recovery (%)	RSD (%)
Estrone (E1)	111,8	5,0	101,3	0,6
17 $\alpha$ -Estradiol (aE2)	132,8	5,7	98,5	6,5
17 $\beta$ -Ethinylestradiol (bE2)	110,5	6,3	97,9	0,6
Estriol (E3)	118,4	6,9	101,0	5,9
17 $\alpha$ -Ethinylestradiol (EE2)	110,9	4,6	98,3	4,1

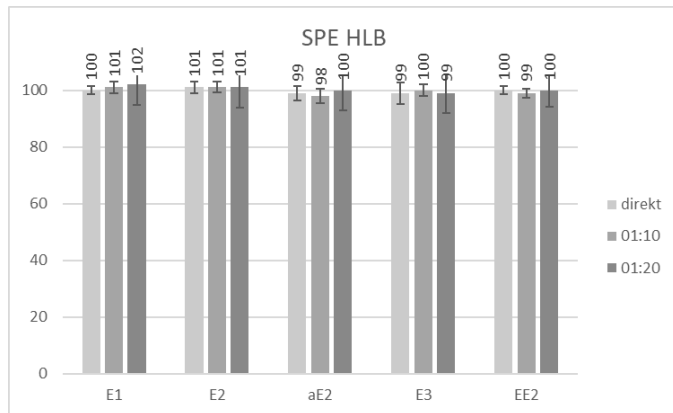
- Sufficient absolute recoveries for E1, E2, alphaE2, EE2 and E3
- Ideal relative recoveries by isotope dilution calibration
- But: no SPM in the water matrix, which can cause clogging of the cartridge

# High complex matrix (Evian water with moderate DOC and with SPM)

Abs. recovery



Rel. recovery



- Sufficient absolute recoveries (left) for E1, E2, alphaE2, EE2 and E3
- Matrix will also be preconcentrated; increasing signals by diluting the sample extracts (graph: 1:10 in 1:20)
- Ideal relative recoveries (right) by isotope dilution calibration

# SPE disk (e.g. combination of filtration and SPE)

- Good compromise of filtration and enrichment
- e.g., HLB, DVB or C<sub>18</sub> disks with 47 mm diameter are feasible (capacity is only defined as L, H and M – not correlation to distinct amounts of SPE sorbents possible)
- Need of special manifold suitable for SPE disks
- Sufficient absolute recovery rates within 80 to 90% in complex water matrix (DOC, SPM and inorganic content)
- Applicable for SPM content higher than 50 mg L<sup>-1</sup> e.g., 500 mg L<sup>-1</sup>
- Preconcentration of DOC content comparable to common SPE



# A typical SPE disk procedure



## 1000 mL water sample

- Spiked with IS-mix at desired level
- 15 min on horizontal shaker
- SPE disk e.g., Atlantic HLB M/L or H



## Conditioning / sample load

- Conditioning with 20 mL ACN
- Conditioning with 3 x 10 mL H<sub>2</sub>O
- Load 1000 mL water sample in approx. 30 min



## Washing step and elution

- Washing cartridge with 10 mL H<sub>2</sub>O
- Drying of SPE disk by applying vacuum for 5 min
- Collect 5 x 10 mL fractions using ACN



## Concentration and analysis

- Concentrate organic fraction to 1 mL (TurboVap)
- Yellowish extract is ready for analysis by LC-MS/MS



## SPE disk

- Different matrix compositions were used:
  - **Low complexity matrix** (Evian water with low DOC and without SPM)
  - **High complexity matrix** (Evian water with moderate DOC and with SPM)
- Relative and absolute recovery rates were determined to compare the different experiments with regards to their performance (applicability in routine analysis)



# SPE disk (combination of filtration and SPE)

**Low complexity matrix** (Evian water with low DOC and without SPM)

Extraction method	Atlantic® C-18 Disks, 47 mm, Evian without additives (1000 mL) and estrogens at 10 ng/L level; no sample clean-up or derivatization			
	Absolute recovery (%)	RSD (%)	Relative recovery (%)	RSD (%)
Parameter				
Estrone (E1)	77,2	12,6	90,0	0,3
17 $\alpha$ -Estradiol (aE2)	NA	NA	86,8	0,4
17 $\beta$ -Ethinylestradiol (bE2)	88,1	16,3	84,5	10,0
Estriol (E3)	82,8	9,8	88,1	1,4
17 $\alpha$ -Ethinylestradiol (EE2)	72,6	10,2	93,8	5,6

- Sufficient absolute recoveries for E1, E2, alphaE2, EE2 and E3
- Ideal relative recoveries by isotope dilution calibration

# SPE disk (combination of filtration and SPE)

High complexity matrix (Evian water with moderate DOC and with SPM)

Extraction method  Parameter	Atlantic® DVB Disks, 47 mm , Evian water (500ml) + 25 mg SPM + DOC (7 mg/L) and estrogens at 5 ng/L level			
	Absolute recovery (%)	RSD (%)	Relative recovery (%)	RSD (%)
Estrone (E1)	92,6	4,0	95,0	7,5
17 $\alpha$ -Estradiol (aE2)	92,4	8,8	88,1	6,5
17 $\beta$ -Ethinylestradiol (bE2)	90,8	9,7	88,1	1,4
Estriol (E3)	99,6	4,4	92,5	4,2
17 $\alpha$ -Ethinylestradiol (EE2)	97,1	10,7	99,0	2,6

- Sufficient absolute recoveries for E1, E2, alphaE2, EE2 and E3
- Sufficient/Ideal relative recoveries by isotope dilution calibration

# SPE disk (combination of filtration and SPE)

**High complex matrix:** unfiltered surface water

Extraction method	Atlantic® C-18 Disks, 47 mm, unfiltered surface water (1000 mL) and estrogens at 10 ng/L level; no sample clean-up or derivatization			
	Absolute recovery (%)	RSD (%)	Relative recovery (%)	RSD (%)
Parameter				
Estrone (E1)	25,3	6,0	90,8	4,3
17 $\alpha$ -Estradiol (aE2)	NA	NA	78,4	9,7
17 $\beta$ -Ethinylestradiol (bE2)	29,4	4,8	90,5	5,0
Estriol (E3)	23,4	10,4	87,8	0,6
17 $\alpha$ -Ethinylestradiol (EE2)	20,7	4,4	104,5	2,6

- Poor absolute recoveries for E1, E2, alphaE2, EE2 and E3
- Sufficient/ideal relative recoveries by isotope dilution calibration

# Mi-SPE (Molecular imprinted polymers)

## PROTOCOL OF PURIFICATION

### Sample preparation

100mL of tap water spiked with 17 $\beta$ -E2-d<sub>3</sub> to a final concentration of 75ng/L was the loading solution.

Purification with a 3mL/100mg AFFINIMIP® SPE Estrogens cartridge

### Equilibration

- 3mL Acetonitrile
- 3mL Water

### Loading solution from sample preparation

#### Washing of interferents

- 3mL water
- 3mL Water/Acetonitrile (60/40)

### Elution (E)

3mL Methanol

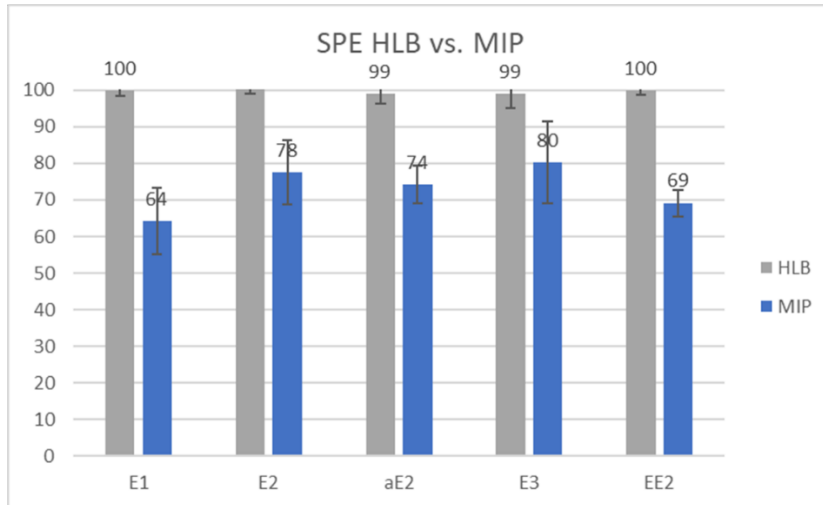
## Publications

Data extracted from **Determination of steroidal oestrogens in tap water samples using solid-phase extraction on a molecularly imprinted polymer sorbent and quantification with gas chromatography-mass spectrometry (GC-MS)**, D. Zacs, I. Perkons, V. Bartkevcis, *Environ Monit Assess* 188, 433, 2016.

- Given protocol by the cartridge provider – limited sample volume of 100 mL – not suitable for estrogens with regards to EU-WFD
- Advantage: specific preconcentration of estrogens – key lock mode of action
- Ideal procedure to separate matrix (e.g., DOC, inorganics, ....) from estrogens (no significant ion suppression)
- Potential: implementation as purification procedure for extracts obtained from preconcentration steps

# Mi-SPE (Molecular imprinted polymers)

Low complexity matrix (Evian water with low DOC and without SPM)



- Mi-SPE: Moderate relative recoveries for E1, E2, alphaE2, EE2 and E3 in comparison to common SPE;
- No high complex matrix experiments possible due to clogging of the cartridge

# Overview of preconcentration methods including limitations and restrictions

(O: fulfilling , +: good, ++: excellent, -: poor, --: unsatisfactory)

Extraction method Parameter	DI	SPE		SPE disk	LLE	Mi-SPE
		off-	online			
Preconcentration	--	++	++	++	O	--
Clean-up	--	+	+	+	O	++
Compatible to common solvents	--	+	+	+	-	O
Selectivity	--	++	++	++	O	++
Time / efficiency	++	+	++	++	+	O

## Conclusions on preconcentration procedures (1: without SPM, 2: with SPM)

Parameter \ Extraction method (Typical preconcentration factor)	SPE (1:1000)		SPE disk (1:1000)		MiSPE (1:100)	
	1	2	1	2	1	2
Estrone (E1)	Green	Green	Green	Green	Yellow	Red
17 $\alpha$ -Estradiol (aE2)	Green	Green	Green	Green	Yellow	Red
17 $\beta$ -Ethinylestradiol (bE2)	Green	Green	Green	Green	Yellow	Red
Estriol (E3)	Green	Green	Green	Green	Yellow	Red
17 $\alpha$ -Ethinylestradiol (EE2)	Green	Green	Green	Green	Yellow	Red

# Conclusions

- SPE and SPE disks: extraction method of choice
- **SPE is applicable for whole water samples with a SPM load less than 50 mg L<sup>-1</sup> to avoid clogging of the cartridge**
- **Higher SPM load: SPE disk is the method of choice**
- Using e.g., HLB sorbent materials (SPE or SPE disk) the DOC will be co-preconcentrated, only 50 to 70% of the initial DOC can be removed by the preconcentration
- This can interfere the analytical method e.g., in case of LC-ESI-MS ion suppression can occur (loss of sensitivity )
- **A further purification of the extract from the preconcentration procedure is recommended by the project consortium**



# Overview on purification/ clean up procedures

## Mi-SPE (molecular imprinted polymers)

### PROTOCOL OF PURIFICATION

#### Sample preparation

100mL of tap water spiked with 17 $\beta$ -E2-d<sub>3</sub> to a final concentration of 75ng/L was the loading solution.

Purification with a 3mL/100mg AFFINIMIP<sup>®</sup> SPE Estrogens cartridge

#### Equilibration

- 3mL Acetonitrile
- 3mL Water

#### Loading solution from sample preparation

#### Washing of interferents

- 3mL water
- 3mL Water/Acetonitrile (60/40)

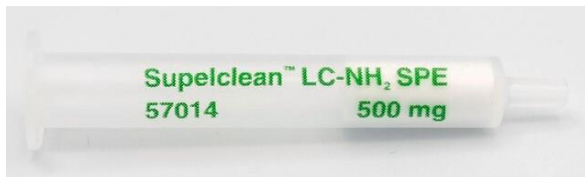
#### Elution (E)

3mL Methanol

- Extract from preconcentration step can be evaporated to at least 1 mL and can be diluted using MilliQ water up to 100 mL
- The volume is compatible to the max volume given by the provider SOP
- Advantage: specific preconcentration of estrogens – key lock mode of action
- Ideal procedure to separate matrix (e.g., DOC, inorganics, ....) from estrogens
- DOC in the Mi-SPE is decreased to a minimum (color changed from yellowish to a complete clear and colorless extract (evaluated by complementary DOC measurements) with sufficient recoveries

## Other SPE cartridges for purification of sample extracts

- For an alternative clean up of the sample extracts a **Supelclean™ LC-NH<sub>2</sub>** SPE (500 mg, 6 mL) cartridge (Merck, Darmstadt, Germany) can be used
- Silica gel based material with amino bounding functional groups
- Increase of matrix components but not as specific as MiSPE



- Also mixed phase cartridges containing e.g., C<sub>18</sub> and aminopropyl functionality are available (e.g., Chromabond NH<sub>2</sub>/C18). Due to limited capacity not sufficient as a two step procedure.

# Overview on purification/ clean up procedures

## Other SPE cartridges for purification of sample extracts



1 mL sample extract

- SPE e.g., Supelclean LC-NH2 (500 mg/6 mL)



Supelclean™ LC-NH, SPE  
57014 500 mg

Conditioning / sample load

- Conditioning with 4 mL MeOH
- Load sample extract



Supelclean™ LC-NH, SPE  
57014 500 mg

Washing step and elution

- Collect 2 mL fraction using MeOH



Concentration and analysis

- Concentrate organic fraction to desired volume (TurboVap)
- Clear extract is ready for analysis by LC-MS/MS

## Two-step procedure preconcentration/purification

- **High complex matrix** (Evian water with moderate DOC and with SPM)
- Relative and absolute recovery rates were determined to compare the different experiments with regards to their performance (applicability in routine analysis)

### TESTED:

1. SPE preconcentration + SPE purification: LC-NH<sub>2</sub>
2. SPE disk preconcentration + SPE purification: LC-NH<sub>2</sub>
3. SPE disk preconcentration + SPE purification: MiSPE

# 1. SPE preconcentration + SPE purification: LC-NH<sub>2</sub>

**High complex matrix** (Evian water with moderate DOC and with SPM)

Extraction method  Parameter	SPE extraction OASIS <b>HLB 500mg/6cc</b> + SPE purification <b>LC-NH2 500mg/3cc, 200 mL</b> Evian water + SPM (50mg/L) + DOC (7 mg/L) and estrogens at <b>0,4 ng/L level excepted 17<math>\alpha</math>-ethynylestradiol at 0,035ng/L</b> . Sample acidification at pH = 5, 1% MeOH added.			
	absolute recovery in %	RSD in %	Isotope dilution recovery in %	RSD in %
Estrone (E1)			98	2
17 $\alpha$ -Estradiol (aE2)			98	15
17 $\beta$ -Estradiol (bE2)			108	11
Estriol (E3)			116	7
17 $\alpha$ -Ethinylestradiol (EE2)			109	10
Estrone-13C3 (E1-13C3)	83	7		
17 $\alpha$ -Estradiol-d2 (aE2-d2)	74	4		
17 $\beta$ -Estradiol-13C3 (bE2-13C3)	81	6		
Estriol-d2 (E3-d2)	80	8		
17 $\alpha$ -Ethinylestradiol-d4 (EE2-d4)	79	5		

## 2. SPE disk preconcentration + SPE purification: LC-NH<sub>2</sub>

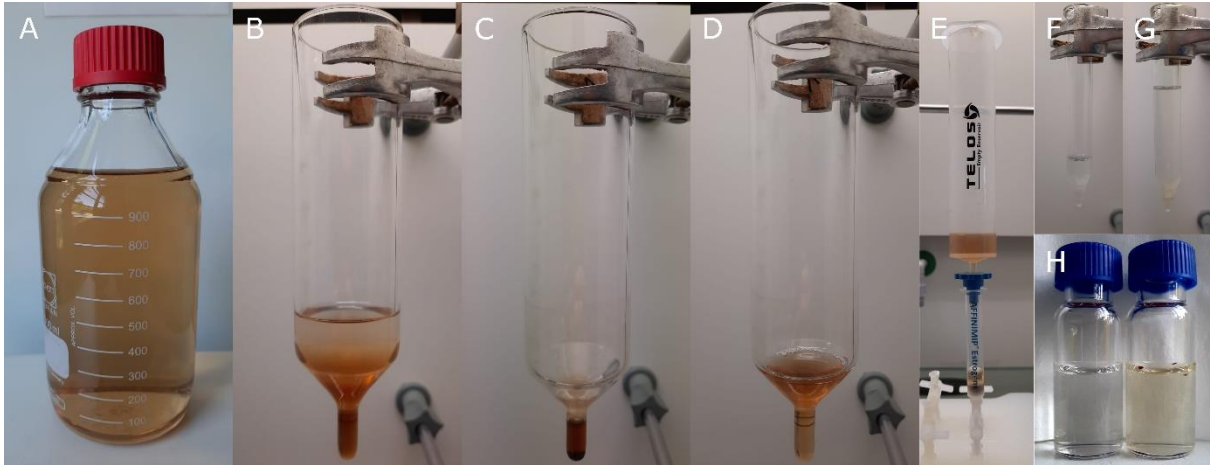
High complex matrix (Evian water with moderate DOC and with SPM)

	Estrogen	Evian <sup>®</sup> + SPM (50 mg L <sup>-1</sup> )		Evian <sup>®</sup> + DOC (5 mg L <sup>-1</sup> )		Evian <sup>®</sup> + DOC (5 mg L <sup>-1</sup> ) + SPM (150 mg L <sup>-1</sup> )	
Abs. rec.	EE2	68 ± 22%		67 ± 3%		88 ± 1%	
	E3	64 ± 14%		62 ± 6%		76 ± 4%	
	aE2	68 ± 20%		70 ± 2%		85 ± 2%	
	bE2	67 ± 19%		69 ± 9%		90 ± 2%	
	E1	68 ± 22%		66 ± 1%		91 ± 4%	
	Rel. rec.	Estrogen	Evian <sup>®</sup> + SPM (50 mg L <sup>-1</sup> ) (%)		Evian <sup>®</sup> + DOC (5 mg L <sup>-1</sup> ) (%)		Evian <sup>®</sup> +DOC (5 mg L <sup>-1</sup> ) - SPM (150 mg L <sup>-1</sup> ) (%)
EE2		109	5	93	5	84	1
E3		97	8	90	9	86	6
aE2		98	1	99	4	98	0,40
bE2		96	2	93	4	92	4
E1		105	1	98	2	96	0,50

## 2. SPE disk preconcentration + SPE purification: MiSPE

High complex matrix (Evian water with moderate DOC and with SPM)

A: START



H left:  
FINAL

- DOC content of the preconcentrated and purified sample is less than 1% than the initial one (A: starting, H left : final)

### 3. SPE disk preconcentration + SPE purification: MiSPE

**High complex matrix** (Evian water with moderate DOC and with SPM)

1 L Evian + 7 mg L<sup>-1</sup> DOC and 50 mg SPM spiked with estrogens at 0.1 ng L<sup>-1</sup>

	HLB disk		MiSPE		HLB disk + MiSPE	
	Relative recovery [%]	Standard deviation [%]	Relative recovery [%]	Standard deviation [%]	Relative recovery [%]	Standard deviation [%]
Estrone	97.00	0.25	92.54	0.105	96.20	0.30
Estradiol	100.02	0.01	100.02	0.01	100.01	0.01
17 $\alpha$ -Estradiol	100.09	0.02	100.07	0.04	100.14	0.06
Estriol	99.65	0.09	99.16	0.21	99.60	0.07
Ethinylestradiol	100.17	0.05	100.21	0.06	100.12	0.04



## Take home message on sample preparation

High complexity matrix:

Preconcentration (SPE, SPE-disks) + purification (SPE, MiSPE)

## Take home message on sample preparation

High complexity matrix:

Preconcentration (SPE, SPE-disks) + purification (SPE, MiSPE)

Thank you for your attention!

